

Biomarkers and their impact on Temporomandibular Disorder: A narrative review

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ABSTRACT

Temporomandibular Disorders encompass a group of musculoskeletal and neuromuscular conditions affecting the temporomandibular joint and associated structures, leading to pain, functional limitations and psychological distress. The variety of TMD symptoms leads to several diagnostic challenges, which can be illustrated by difficulties in treatment planning. Biomarkers are biological molecule found in blood, other body fluids, or tissues that is a sign of a normal or abnormal process, or of a condition or disease. They enhance our ability to identify relationships between environmental chemicals and human diseases, thereby improving our capacity to diagnose, monitor, or predict disease risk. They can serve as an early warning systems for our health. They are often measured and evaluated using blood, synovial fluid, saliva, and other bodily fluids to examine normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention. These biomarkers are a relatively new clinical toolset categorised by their clinical applications. This article presents a review on the impact of various biomarkers on TMD.

Keywords: TMD, Biomarkers, Diagnosis

INTRODUCTION

Biomarkers are objective measurements that capture the current state of an organism or cell. They enhance our ability to identify relationships between environmental chemicals and human diseases, thereby improving our capacity to diagnose, monitor, or predict disease risk. They can serve as an early warning systems for our health. Biomarkers are often measured and evaluated using blood, synovial fluid, saliva, and other bodily fluids to examine normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention.¹ These biomarkers are a relatively new clinical toolset categorized by their clinical applications. The four main classes are molecular, physiologic, histologic and radiographic biomarkers. All four types of biomarkers have a clinical role in narrowing or guiding treatment decisions and follow a sub-categorisation of being either predictive, prognostic, or diagnostic.²

Pathophysiology & Diagnostic challenges in TMD

Temporomandibular disorder (TMD) is a musculoskeletal disorder that is manifested through continuous pain in the temporomandibular joint (TMJ), masticatory muscle, and the periauricular region.³ Meanwhile, the most common non-odontogenic orofacial pain is a pain in the temporomandibular area.^{4,5} Other related symptoms like hyoid bone tenderness, abnormal swallowing, and tinnitus have an impact on an individual's sleep, quality of life, and psychological well-being.⁶ Therefore, these symptoms also result in depression, stress, anxiety, adverse impacts on energy level, emotional condition and social function.^{7,8} The occurrence of TMD symptoms ranges from 21.5% to 50.5%, with women

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How to cite the article: Philip G B, Murupel A M, P Jayanthi, T Nripan. Biomarkers and their impact on Temporomandibular Disorder: A narrative review. Oral MaxillofacPathol J 2026; 17(2); 308-317.

Source of Support: Nil

Conflict of Interest: None

experiencing more than men⁶. Constitutional, hormonal, psychological, biological, anatomical, and behavioural factors are the causes of this discrepancy.^{9,10,11} The pathophysiology of TMD is multifactorial, which includes biopsychosocial, neuromuscular, biomechanical, and biological factors.^{6,12}

The variety of TMD symptoms leads to several diagnostic challenges, which can be illustrated by difficulties in treatment planning.¹³ In most cases, diagnosis is based on history physical & radiographic examination.¹⁴ Orthopantomogram (OPG), Lateral Cephalogram, Computed Tomography (CT) are commonly used. However to become radiographically evident considerable amount of bone loss around 30% should happen.¹⁵ Therefore, some new diagnostic approaches should be considered which might help in early detection of bone disorders.

Diagnostic Biofluids : From Synovial fluids to Saliva

Biomarkers are analyzed using biological samples such as tissue biopsy and biofluid plasma, saliva, synovial fluid, and cerebrospinal fluid. Previously, blood serum was the gold standard for biomarker assessment and validation, and this was followed by synovial fluid. Joint specific biochemical indicators cannot enter the bloodstream because the synovium, the semi-permeable barrier lining the synovial joint naturally blocks the passage of large molecules. Hence, synovial fluid analysis can uniquely pinpoint joint conditions, providing real-time information.¹⁶ One study unveiled a correlation between specific TMD biomarkers and morphological variations in distinct anatomical regions of the TMJ condylar surface and stated that the condyle's front surface had shown bone apposition and repair, while regions of bone resorption were specifically observed near the lateral pole.¹⁷

In recent years, there has been growing interest in exploring the potential role of salivary biomarkers in TMDs.^{18,19,20} Saliva, as a readily accessible biofluid, offers numerous advantages as a diagnostic medium, such as noninvasiveness, cost-effectiveness and ease of collection. Furthermore, saliva plays a critical role in maintaining a protective barrier over oral tissues, which helps mitigate mechanical stress and friction during mastication.^{24,25,26,27,28}

Saliva is also essential for lubricating the temporomandibular joint (TMJ) during movement. Changes in saliva's pH and buffering ability can alter the oral environment.³⁴ Acidic surroundings can cause enamel demineralisation, which can exacerbate symptoms of TMD.³⁵ Even a decrease in salivary flow or a change in the content of the saliva could cause insufficient lubrication, which would increase friction and damage the TMJ structures.³³

Salivary biomarkers, such as proteins, enzymes, inflammatory mediators and genetic markers, hold promise as objective indicators of disease presence, progression and treatment outcomes in TMDs.^{21,22,23} According to new research, the pathophysiology of painful

TMD might be due to inflammation or the release of inflammatory cytokines, possibly due to mechanical overload, local hypoxia, microtrauma, and reduced adaptive capacity of cartilage.¹⁶ By examining the molecular signatures present in saliva, researchers aim to unravel the complex pathophysiological processes associated with TMDs and identify novel diagnostic and therapeutic targets. In this context, saliva has emerged as a promising medium for obtaining a real-time biomarker.^{29,30,31,32,36,37}

A list of various biomarkers and their consequent actions on TMD has been summarized in Table 1. Accordingly, this article aims to provide a comprehensive review of recent updates on TMD biomarkers, focusing on their diagnostic utility as well as their promising potential for suppressing inflammation and eliminating pain

Molecular Profiling of TMD Biomarkers

4.1 Prostaglandins

They are synthesized via the cyclooxygenase (COX) enzymatic pathway during acute and chronic tissue stress, PGE2 is a primary mediator of inflammatory hyperalgesia.¹⁶ Elevated

levels of PGE2 in the synovial fluid of TMD patients correlate significantly with reported TMJ pain during mandibular movement.³⁸ This correlation underlines its clinical value as a functional marker of localized joint inflammation.

4.2 Cytokines

They are the essential polypeptide mediators of critical and severe inflammation.³⁹ These molecules function as complex immunological networks containing pro-inflammatory cytokines, including interleukin-1 (IL-1), interleukin-6 (IL-6), and tumor necrosis factor (TNF), and anti-inflammatory mediators, such as interleukin -10 (IL-10) and transforming growth factor-beta (TGF- β). The balance between them dictates either joint homeostasis or progressive degradation. The high concentrations of pro-inflammatory cytokines are strongly linked to osteoarthritis (OA) and internal derangement (ID).⁴⁰ These mediators contribute to the degradation of cartilage and bone joints by releasing proteinases and other inflammatory molecules.

- **Interleukin 1 (IL-1):** The IL-1 family includes the pro-inflammatory ligands IL-1 α and IL-1 β , balanced by their competitive receptor antagonist, IL-1ra.⁴¹ An imbalance favoring elevated synovial IL-1 α and IL-1 β concentrations disrupts TMJ metabolic homeostasis, triggering the development and accelerating the severity of structural TMD.⁴²
- **Interleukin 6 (IL- 6):** IL-6 is a circulating cytokine produced by cells (endothelial cells, adipose tissue, T-cells, smooth muscle cells, and macrophages).⁴³ It regulates oncogenesis, hematopoiesis, and immune responses.⁴⁴ Crucially, it acts as a primary molecular bridge driving the transition from acute to chronic inflammation by inducing osteoclast progenitor differentiation and activating localized bone resorption.⁴⁵ Following acute tissue injury, systemic plasma concentrations of IL-6 rise within 60 minutes, peaking between 4 to 6 hours and persisting for up to 10 days.^{46,47} Due to its broad systemic involvement in hematopoiesis and oncogenesis, isolating an IL-6 reading to confirm a specific diagnosis of TMD requires localized synovial fluid sampling, which is heavily limited by clinician technique and patient discomfort
- **Interleukin 8 (IL- 8):** IL-8 is produced by macrophages, endothelial cells, and epithelial tissues.⁴⁸ It induces targeted chemotaxis and recruits neutrophils into the synovial fluid,⁴⁹ exacerbating active joint inflammation especially in angiogenic diseases, including rheumatoid arthritis (RA).^{50,51} This chemokine is highly sensitive to local tissue hypoxia,⁵² and its presence has been verified at the posterior disc attachment in patients diagnosed with internal derangement.⁵³ It also leads to increase in the amount of inflammatory cells in TMJ.^{54,55} Because IL-8 is heavily dependent on localized hypoxia as a regulatory trigger, its presence is highly variable. It may drop below detectable diagnostic thresholds in chronic, non-inflammatory myofascial TMD or stabilized disc displacements where active tissue strain has subsided.



Table 1 Biomarkers and their actions

Thamer, S.R. and Diajil, A.R. (2024) ⁹⁸	Saliva	MMPs	MMP-2 and MMP-9 positively correlated with pain and joint click and negatively with mouth opening.
AlSahman, L. et al. (2024) ⁹⁹	Saliva	Cortisol	↑cortisol: Disc Displacement without Reduction
Ismah, N. et al. (2024) ¹⁰⁰	Saliva	IL-1β	Pain-related or joint TMDs
Kim, Y. et al. (2023) ¹⁰¹	Blood	ILs, Cortisol	↑IL-1β, ↑IL-4, ↑IL-8, and ↑IL-17 showed sufficient strength
Ornek et al. (2023) ¹⁰²	Saliva	Cortisol, IL-1β, TNF-α and MMP3	Although not significantly greater, cortisol and depression/anxiety were higher in the study group. Additionally, there was no discernible difference in the others
Shao, B. et al. (2023) ¹⁰³	Synovial fluid	ILs, PGE2	High levels of biomarkers, Osteoarthritis > Disc Displacement
Aparna, N. et al. (2023) ¹⁰⁴	Saliva	Cortisol	No statistically significant difference in salivary cortisol level between cases and controls.
Kazan, D. et al. (2023) ¹⁰⁵	Saliva, blood	IL-6,	Strong positive correlation between pain
Zwiri, A.M. et al. (2022) ¹⁰⁶	Blood	ILs	IL-8 may serve as a potential biomarker for TMJ pain:
Ulmner, M. et al. (2022) ¹⁰⁷	Synovial fluid	ILs, TNF-α	IL-1β and TNF-α were significantly associated with TMJ palpation pain. TNF-α also correlated with subjective TMJ pain. IL-1β was linked to synovitis, which contributes to pain
Bayındır, S., et al. (2022) ¹⁰⁸	Synovial fluid	PGE2	PGE2 are linked to localized TMJ pain and are elevated in joints with degenerative changes
Venkatesh, S.B. et al. B. et al. (2021) ¹⁰⁹	Saliva	Cortisol	Cortisol: strong association with stress and TMD severity.
Son, C. et al. (2021) ¹¹⁰	Blood	ILs, TNF-α, PGE2,	TMD—higher pain intensity/duration and ↑IL-8 and IgG levels link chronic pain and systemic inflammation.
Ulmner, M. et al. (2020) ¹¹¹	Synovial fluid	ILs	Disc Displacement without Reduction: IL-1↑ (female > male)
Loreto, C. et al. (2020) ¹¹²	Synovial fluid	MMPs	MMP-7 and MMP-9 overexpressed in Disc Displacement without Reduction
Khayam et al. (2020) ¹¹³	Saliva	Cortisol	In comparison to the Control group, the TMD group had considerably higher levels of cortisol.
Shoukri et al. (2019) ¹¹⁴	Saliva	MMP3, PAI-1 IL1α, IL6, MMP 3, MMP7, PAI-1, TNFα	MMP- 3, and PAI- 1 were positively expressed and significantly correlated to condylar variance in TMD patients among all the biomarkers evaluated.
Xiong et al. (2019) ¹¹⁵	Synovial fluid	IL6	Elevated leptin levels in temporomandibular joint promote proinflammatory cytokine IL-6 expression in synovial fibroblasts
Yang et al. (2019) ¹¹⁶	Synovial fluid	IL8	MRI- levels of cytokines (IL-8, and TNF) in the synovial fluid were increased in TMD cases
Ganti, S. et al. (2018) ¹¹⁷	Synovial tissue	ILs, TNF-α, PGE2	IL-1β, TNF-α, PGE2, and IL-6- TMD
Ce et al. (2018) ¹¹⁸	Saliva	IL-1β	Regardless of a fibromyalgia assessment, the patients with TMD alone had significantly higher IL-1 levels, whereas the group with fibromyalgia alone did not differ significantly from the Control Group
Watanabe et al., (2017) ¹¹⁹	Synovial fluid	IL-1β, IL-6, and IL-8	Protein concentrations of IL-1, IL-6, and IL-8 ↑ - ELISA Test- inflammatory condition in the TMJ.
Kobayashi et al (2017) ¹²⁰	Saliva	Cortisol	Cortisol levels were assessed to not be significantly linked to TMD, although anxiety related symptoms were more severe in the TMD group

Dawson et al (2016) ¹²¹	Synovial fluid	Dopamine	In myofascial pain and degenerative diseases, dopamine might be involved in modulating peripheral pain
Ahmed et al. (2015) ¹²²	Synovial fluid	TNF	TMJ pain -increased inflammatory activity
Lin, S.L., et al. (2015) ¹²³	Blood	Cortisol	↑Cortisol in Disc Displacement without Reduction: clinical indicator for distinguishing disc displacement disorders
Cevidane, L.H. et al. H. et al. (2014) ¹²⁴	Synovial fluid and blood	MMPs, ILs, TNF	Osteo Arthritis showed bone resorption: MMPs linked to bone apposition, while IL-6 and TNFα linked to bone resorption.
Vos, L.M. et al. (2013) ¹²⁵	Synovial fluid	IL-1β, TNF-α, PGE2	High collagen-II levels suggest it may be a useful marker for cartilage degradation.
Wake et al. (2013) ¹²⁶	Synovial fluid	IL-6	Concentrations of aggrecan, IL-6, and VEGF-A were significantly higher in the group with synovial chondromatosis of TMJ
Nogami et al. (2013) ¹²⁷	Synovial fluid	IL-6	Concentration of IL-6 were significantly higher in patients with joint effusion
Kim et al. (2012) ¹²⁸	Synovial fluid	IL-1β, IL-2, IL-6, IL-8, IL-10, and TNF-α	interleukin (IL)-1β, IL-2, IL-6, IL-8, IL-10 and tumour necrosis factor (TNF)-α were detected in the TMD
Slade, G.D. et al. D. et al. (2011). ¹²⁹	Blood	ILs	Localized TMD linked to IL-1 and widespread TMD linked to IL-8.
Miller, K. E et al (2011). ⁷⁸	Synovial fluid	Glutamate	Involved in pain processing, peripheral nociception, and central sensitization of painful TMD
Alstergren, P et al. (2010) ⁸⁰	Synovial fluid	Glutamate	Glutamate evokes immediate pain in the healthy human TMJ that is partly mediated by peripheral NMDA receptors in the TMJ.
Lee et al. (2010) ¹³⁰	Synovial fluid	IL-6 and TNF-α	Elevation of TNF-alpha and IL-6, in the synovial fluid of temporomandibular disorder patients with symptoms of pain, mouth opening limitation, and clicking.
Hamada et al. (2008) ¹³¹	Synovial fluid	IL-6, IL-8 & IL10	Presence of IL-10 is a significant predictor of successful outcome of TMJ irrigation for Chronic Closed Lock
Vernal et al. (2008) ¹³²	Synovial fluid	IL-1b, IL-2, IL-12, TNF-α and TNF-β	IL1β, TNF-α, IL10, IL12, and IL17 are involved in the OA-TMJ pathogenesis
Sato, J. et al. (2007) ⁵⁴	Synovial fluid	IL 8	↑IL-8 in TMD, no significant link to pain or inflammation severity
Yoshida, K. et al. (2006) ⁶⁸	Synovial fluid	MMPs	↑MMP-9 in severe TMJ Osteoarthritis and disc displacement. ↑MMP-2 elevated in early Osteoarthritis.
Kardel et al (2006) ¹³³	Synovial fluid	IL-1α, IL-1β	The cytokines (IL-1α, IL-1β) ↑ in TMD
Matsumoto et al., (2006) ¹³⁴	Synovial fluid	IL8	Elevated cytokine levels in TMD
Kaneyama, K. et al. (2004) ¹³⁵	Synovial fluid	ILs	↑IL-6 and ↑IL-11 in joints with condylar bone changes: osseous degeneration.
Nishimura et al., (2004) ¹³⁶	Synovial fluid	IL-1α, IL-1β,	Presence of IL-1β and IL-6 in synovial fluid are indicators for internal derangement of the TMJ.
Kardel, R. et al. (2003) ¹³⁷	Synovial fluid	ILs, TNF-α	Osteoarthritis joints: ↑IL-1α, ↑IL-1β
Nishimura et al., (2002) ⁷⁷	Synovial Fluid	Bradykinin	Involved in TMD pain secondary to disc disorders degenerative joint diseases and muscle soreness
Srinivas, R. et al. ⁷²	Synovial fluid	MMPs	↑MMP-1, ↑MMP-2, ↑MMP-8, ↑MMP-9, ↑MMP-13 in mild TMJ-ID—active collagen degradation
Tanaka, A. et al. (2001) ¹³⁸	Synovial fluid	MMPs	↑MMP-2 and ↑MMP-9 in Disc Displacement without Reduction > Disc Displacement with Reduction—diagnostic markers.

Shinoda et al (2000) ¹³⁹	Synovial fluid	IL6	IL-6 - chronic TMJ disorders
Ernberg et al (2000) ⁸⁹	Synovial Fluid	Serotonin	Involved in Peripheral and central pain mechanism of painful TMD. Sensitize neuropeptides and other neurotransmitters such as glutamate are involved in the modulation and control of TMD pain.
Murakami, K.I. et al. (1998) ¹⁴⁰	Synovial fluid	PGE2	↑PGE2 linked to TMJ degeneration
Kubota, E. et al (1998) ¹⁴¹	Synovial fluid	MMPs and ILs	IL-1β ↑Disc Displacement without Reduction, IL-1β ↑Osteoarthritis, IL-6 ↑Osteoarthritis —markers linked to cartilage degradation and pain in TMD
Takahashi et al. (1998) ¹⁴²	Synovial Fluid	IL-1β, and TNF-α	Increased levels of proinflammatory cytokines IL-1β in TMD ,
Kubota, E. et al. (1997) ¹⁴³	Synovial fluid	IL-1β, MMP, TNF-α	↑IL-1β in TMD (osteolytic changes TMJs) ↑MMP-3 linked to cartilage degradation —early markers of TMJ deterioration and pain.
Fu et al. (1995) ¹⁴⁴	Synovial Fluid	TNF-α & IL6	Increased in TMD

- Tumour Necrosis Factor (TNF):** TNF is regarded as the master pro-inflammatory cytokine.⁵⁶ It is present in the form of membrane TNF (mTNF), pro-TNF and soluble TNF (sTNF).^{57,58} Specifically, mTNF is a transmembrane protein which is transformed into sTNF when divided by TNF-converting enzyme (TACE).^{57,59} Furthermore, TNF plays an essential role in leukocyte recruitment, monocyte chemo-attraction, development of apoptosis, and enhanced management of adhesion molecule expression.⁶⁰ Immune cells, such as natural killer (NK) cells, activated macrophages/monocytes and activated T cells, and other non-immune cells, such as fibroblasts and endothelial cells, represent TNF.⁶¹ TNF instigates a broad inflammatory cascade by driving the subsequent transcription of downstream mediators like IL-1 and IL-6.^{62,63} Clinically, synovial TNF concentrations serve as a valuable predictive biomarker.⁶⁴ Another study examined the role of TNF in synovial fluid as a predictor of the therapeutic effect of intra-articular glucocorticoid injection on TMJ in TMD patients and a positive correlation was found in patients with TMD and TNF with chronic inflammatory connective tissue disease.⁶⁵ Consequently, a clinician cannot use a positive TNF reading to conclusively diagnose primary TMD unless comprehensive systemic rheumatologic screenings are completed to rule out broader cross-reactive conditions.

4.3 Proteinases

These enzymes hydrolyse peptide links to break down proteins. Matrix Metalloproteinases (MMPs) are tissue-degrading proteolytic enzymes deeply involved in TMJ inflammation and pain.⁶⁶ Research shows that joint inflammation upregulates MMP-1, MMP-2, and MMP-13, accelerating degenerative changes and pain progression.⁶⁷⁻⁶⁹ In patients with disc disorders

and joint degeneration, TMJ synovial fluid contains elevated levels of collagenases (MMP-1, MMP-8, MMP-9, MMP-13), stromelysin (MMP-3), and gelatinases (MMP-2, MMP-7).⁷⁰⁻⁷⁴ This expression is also influenced by psychological stress, which elevates MMP-3 levels in correlation with TMD severity.⁷⁵ Furthermore, MMP-2 and MMP-9 have been detected in the trigeminal ganglion during TMJ inflammation.⁷⁶ Ultimately, these enzymes serve as viable therapeutic targets for TMD pain management.

4.4 Neuropeptides

These are small protein-like molecules, specifically short chains of amino acids, that act as signalling molecules in the nervous system and other tissues. They are synthesized and released by neurons and play a crucial role in various physiological processes like pain perception, stress response, etc. Bradykinins are mediators of inflammation, which are crucial for nociception and sensitisation. It is a powerful vasodilator and promoter of vascular permeability, elevated synovial bradykinin levels correlate positively with verified degrees of joint inflammation in patients with structural disc disorders.⁷⁷ This provides excellent diagnostic sensitivity for identifying active internal joint derangements.

4.5 Neurotransmitters

These are chemical messengers that carry messages from neurons to other cells, such as muscles or glands, across synapses. Neurotransmitters are important for both central processing and peripheral or central sensitisation in painful TMDs. They have a significant influence on mood and behavior in addition to pain processing.

- Glutamate :** Glutamate acts as the primary excitatory neurotransmitter within afferent sensory nerves, it relays peripheral pain data from the TMJ and trigeminal ganglion straight to the central nervous system

(CNS).²² Glutamate is highly sensitive to painful TMD states, accumulating significantly in the synovial fluid of joints undergoing advanced degenerative diseases secondary to rheumatoid arthritis.^{78,79,80} However, because glutamate can easily leak into synovial tissues directly from the plasma during systemic excitatory states, its absolute diagnostic specificity for primary isolated joint pathology can be compromised limiting its use to specialized diagnostic centers.

- **Dopamine:** Dopamine participates in a complex regulatory balance governing motor control, cognition, and nociceptive processing across both the central and peripheral nervous systems. In patients presenting with myofascial pain TMD (M-TMD), peripheral plasma dopamine levels rise significantly⁸¹ and correlate tightly with masticatory muscle pain intensity and elevated levels of perceived mental stress.^{82,83} Plasma dopamine acts as a sensitive index of patients overall psychological distress and systemic sympathetic nervous system activation. It exhibits high sensitivity for mapping masseter muscle pain intensity.⁸⁴ However, it has very low specificity for structural joint pathology.⁸⁵ Because central dopamine neurotransmission changes during any chronic pain condition, plasma dopamine readings cannot be used to diagnose TMD independently; they must be interpreted alongside a comprehensive physical and psychological evaluation.
- **Serotonin:** Serotonin or 5-hydroxytryptamine (5-HT) modulates physiological processes in both the central and peripheral nervous systems. It is observed that in response to tissue trauma and inflammation, 5-HT₃, a subtype of 5-HT, is released and sensitizes peripheral neurons, which seems to be important for mediating pain from the periphery.^{86,87} Patients with chronic myalgia exhibit an increased density of these serotonin receptors directly on the sensory nerves of the masseter muscle.^{88,89} Serotonin is an exceptionally robust diagnostic biomarker as it is completely undetectable in the TMJ synovial fluid of healthy individuals. Consequently, the mere emergence of serotonin in synovial fluid provides nearly 100% diagnostic specificity for joint pathology. Elevated levels in the synovial fluid, serum, and saliva correlate with mechanical jaw movement pain, chronic myalgia, and localized allodynia.^{87,88,89,90} Furthermore, local and systemic 5-HT levels partially determine the therapeutic success of intra-articular glucocorticoid injections for TMJ pain, making serotonin a highly reliable biomarker for painful TMD.⁹¹ While its presence in synovial fluid is definitive for pathology, its systemic presentation in serum and saliva can be altered by unrelated emotional disorders or medications (such as antidepressants), necessitating careful pharmacological charting during patient intake.

4.6 Salivary Cortisol

Cortisol, known more formally as hydrocortisone, is a steroid hormone, more specifically a glucocorticoid, produced

by the zona fasciculata of the adrenal cortex. Its main actions include suppressing the immune system, promoting the metabolism of fat, protein, and carbohydrates, and raising blood sugar levels through gluconeogenesis. It also decreases bone formation.^{92,93} TMD patients are likely to present higher salivary cortisol levels as a response to the stress of TMD pain.⁹⁴ Furthermore, it has been demonstrated that a greater cortisol level is connected with a higher Visual Analogue Scale (VAS) score.⁹⁵ Additionally, salivary cortisol levels were found to be significantly higher in patients with disc displacement without reduction and limited mouth opening.⁹⁶ Despite its ease of collection and analytical clarity, cortisol secretion follows a strict diurnal rhythm, peaking immediately upon waking and tapering throughout the day. To maintain diagnostic accuracy and avoid false positives or negatives, saliva sampling must be strictly standardized to specific collection times, and clinicians must control for confounding variables such as acute sleep deprivation or systemic steroid therapy.

Future perspectives

The biomarkers help to map a patient's profile by providing an invaluable clinical toolset that addresses traditional diagnostic challenges. These molecular messengers reveal early signs of localized disease, underlying risks, and actual treatment responses. Gene mapping in TMD could be a future diagnostic tool in identifying inherited genetic variant that may cause joint pain & inflammation. This is possible because the TMJ's connective tissues and cartilage uniquely develop from neural crest cells rather than the mesoderm.⁹⁷

CONCLUSION

TMDs are among the most common maxillofacial disorders, despite the various diagnostic procedures involved, the biomarker profile of patients with TMDs serves as a critical diagnostic tool.

The pathophysiology of painful TMD involves a shift from localized inflammation and cartilage degradation to broader neuroendocrine involvement in chronic states. While acute painful TMD is heavily characterized by inflammatory cytokines, prostaglandins, and matrix metalloproteinases driven by mechanical stress, chronic cases are further compounded by alterations in neurotransmitters, neuropeptides, and cortisol levels.

REFERENCE

1. Abbass M, Rady DEI, Moshi S, Ahmed Radwan, I, Wadan A.S, Dorfer C.E, El-Sayed K.M.F. The Temporomandibular Joint and the Human Body: A New Perspective on Cross Talk. *Dent J.* 2024; 12: 357.
2. Califf RM. Biomarker definitions and their applications: *Exp Biol Med* (Maywood). 2018 Feb;243(3):213-221
3. Gil-Martínez A, Paris-Aleman A, López-de-Uralde-Villanueva I, La Touche R. Management of pain in patients with temporomandibular disorder (TMD): Challenges and solutions. *J. Pain Res.* 2018;11: 571.
4. Zakrzewska J.M. Temporomandibular disorders, headaches and chronic pain: *J. Pain Palliat. Care Pharm.* 2015; 29: 61–63.
5. Conti P.C, Pinto-Fiamengui L.M, Cunha C.O, Conti A.C. Orofacial pain and temporomandibular disorders: The impact on oral health and quality of life. *Braz. Oral Res.* 2012; 26: 120–123.
6. Xu G-Z, Jia J, Jin L, Li J-H, Wang Z-Y, Cao D-Y. Low-Level Laser



- Therapy for Temporomandibular Disorders: A Systematic Review with Meta-Analysis. *Pain Res. Manag.* 2018; 1–13.
7. Oliveira L.K, Almeida Gde A, Lelis E.R, Tavares M, Fernandes Neto A.J. Temporomandibular disorder and anxiety, quality of sleep, and quality of life in nursing professionals. *Braz. Oral Res.* 2015; 29:1–7.
 8. Schierz O, John M.T, Reissmann D.R, Mehrstedt M, Szentpetery A. Comparison of perceived oral health in patients with temporomandibular disorders and dental anxiety using oral health-related quality of life profiles. *Qual. Life Res.* 2008; 17: 857–866.
 9. Mohlin B, Axelsson S, Paulin G, Pietila T, Bondemark L, Brattstrom V, et al. TMD in relation to malocclusion and orthodontic treatment. *Angle Orthod.* 2007; 77:542–548.
 10. Poveda Roda R, Bagan J.V, Diaz Fernandez J.M, Hernandez Bazan S, Jimenez Soriano Y. Review of temporomandibular joint pathology. Part I: Classification, epidemiology and risk factors. *Med. Oral Patol. Oral Cirugía Bucal* 2007; 12: E292–E298.
 11. Bagis B, Ayaz E.A, Turgut S, Durkan R, Ozcan M. Gender difference in prevalence of signs and symptoms of temporomandibular joint disorders: A retrospective study on 243 consecutive patients. *Int. J. Med. Sci.* 2012; 9: 539–544. [CrossRef]
 12. Chisnoiu A.M, Picos A.M, Popa S, Chisnoiu P.D, Lascu L, Picos A, et al. Factors involved in the etiology of temporomandibular disorders – A literature review. *Clujul Med.* 2015; 88: 473–478.
 13. Manfredini D, Bucci M.B, Nardini L.G. The diagnostic process for temporomandibular disorders. *Stomatologija* 2007; 9: 35–39.
 14. Gauer R, Semidey M.J. Diagnosis and treatment of temporomandibular disorders. *Am. Fam. Physician.* 2015; 91: 378–386.
 15. Loffler MT, Sollmann N, Mei K, Valentinitich A, Noël PB, Kirschke JS, Baum T. X-ray-based quantitative osteoporosis imaging at the spine. *Osteoporos Int.* 2020 Feb;31(2):233–250
 16. Goulet J. P & Velly A. M (eds). *Orofacial Pain Biomarkers* (Springer, 2017)
 17. Tanaka E, Detamore MS, Mercuri LG. Degenerative disorders of the temporomandibular joint: etiology, diagnosis, and treatment. *J Dent Res.* 2008;87:296–307.
 18. Minervini G, Franco R, Marrapodi MM, Fiorillo L, Cervino G, Cicciù M. Prevalence of temporomandibular disorders (TMD) in pregnancy: a systematic review with meta- analysis. *J Oral Rehabil.* 2023;50:627–634.
 19. Minervini G, Franco R, Marrapodi MM, et al. Correlation between temporomandibular disorders (TMD) and posture evaluated through the diagnostic criteria for temporomandibular disorders (DC/TMD): a systematic review with meta- analysis. *J Clin Med.* 2023;12:2652.
 20. Minervini G, Franco R, Marrapodi MM, Ronsiville V, Shapira I, Cicciù M. Prevalence of temporomandibular disorders in subjects affected by Parkinson disease: a systematic review and meta-analysis. *J Oral Rehabil.* 2023;50:877–885.
 21. Li DTS, Leung YY. Temporomandibular disorders: current concepts and controversies in diagnosis and management. *Diagnostics.* 2021;11:459.
 22. Chen Y, Yao C, Chang Z, et al. Occlusal function and electromyographic activity of masticatory muscles in skeletal class III patients with different patterns of mandibular asymmetry. *J Oral Rehabil.* 2023;50:276–285.
 23. Zhao M, Han M, Habumugisha J, et al. Electromyographic activities of the jaw and facial muscles in subjects with different vertical skeletal patterns and breathing modes. *J Oral Rehabil.* 2023;50:351–359.
 24. Shaikh M, Alnazzawi A, Habib S, Lone M, Zafar M. Therapeutic role of nystatin added to tissue conditioners for treating denture-induced stomatitis: a systematic review. *Prosthesis.* 2021;3:61–74.
 25. Ahmed N, Humayun M, Abbasi M, Jamayet N, Habib S, Zafar M. Comparison of canine guided occlusion with other occlusal schemes in removable complete dentures: a systematic review. *Prosthesis.* 2021;3:85–98.
 26. D’Addazio G, Xhajanka E, Cerone P, et al. Traditional removable partial dentures versus implant- supported removable partial dentures: a retrospective, observational Oral health related quality of life study. *Prosthesis.* 2021;3:361–369.
 27. Pugliese A, Cataneo E, Fortunato L. Construction of a removable partial denture (RPD): comparison between the analog procedure and the selective laser melting procedure. *Prosthesis.* 2021;3:428–436.
 28. Mangone E, Cataneo E, Fortunato L, Cresti L. Functional removable prosthetic rehabilitation using the electronic Condylograph: a case report. *Prosthesis.* 2021;3:437–445.
 29. de Andrade CM, Galvão- Moreira LV, de Oliveira JFF, et al. Salivary biomarkers for caries susceptibility and mental stress in individuals with facial pain. *Cranio.* 2021;39:231–237.
 30. Campus G, Diaz- Betancourt M, Cagetti M, et al. Study protocol for an online questionnaire survey on symptoms/ signs, protective measures, level of awareness and perception regarding COVID 19 outbreak among dentists. A global survey. *Int J Environ Res Public Health.* 2020;17:5598.
 31. Adina S, Dipalma G, Bordea IR, et al. Orthopedic joint stability influences growth and maxillary development: clinical aspects. *J Biol Regul Homeost Agents.* 2020;34:747–756.
 32. Marrelli M, Tatullo M, Dipalma G, Inchingolo F. Oral infection by staphylococcus aureus in patients affected by white sponge nevus: a description of two cases occurred in the same family. *Int J Med Sci.* 2012;9:47–50.
 33. Nitzan DW. The process of lubrication impairment and its involvement in temporomandibular joint disc displacement: a theoretical concept. *J Oral Maxillofac Surg.* 2001;59:36–45.
 34. Chang H, Israel H. Analysis of inflammatory mediators in temporomandibular joint synovial fluid lavage samples of symptomatic patients and asymptomatic controls. *J Oral Maxillofac Surg.* 2005;63:761–765.
 35. Rezende RLS, Bonjardim LR, Neves ELA, et al. Oral health, temporomandibular disorder, and masticatory performance in patients with Charcot- Marie- tooth type 2. *Sci World J.* 2013;2013:1–8.
 36. Ernberg M. The role of molecular pain biomarkers in temporomandibular joint internal derangement. *J. Oral. Rehabil.* 2017; 44:481–491.
 37. Doshi L.T. et al. Biomarkers in temporomandibular disorder and trigeminal neuralgia: a conceptual framework for understanding chronic pain. *Can. J. Pain.* 2020; 4: 1–18
 38. Alstergren P & Kopp S. Prostaglandin E2 in temporomandibular joint synovial fluid and its relation to pain and in ammatory disorders. *J. Oral. Maxillofac. Surg.* 2000;5: 180–188.
 39. Feghali C.A, Wright T.M. Cytokines in acute and chronic inflammation. *Front. Biosci.* 1997; 2: d12–d26.
 40. Campos M.I.G, Campos P.S.F, Line S.R.P. Inflammatory cytokines activity in temporomandibular joint disorders: A review of literature. *Braz. J. Oral Sci.* 2006; 5: 1054–1062.
 41. Garlanda C, Dinarello C.A, Mantovani A. The interleukin-1 family: Back to the future Immunity. 2013; 39:1003–1018
 42. Poluha R.L, Grossmann E. Inflammatory mediators related to arthrogenic temporomandibular dysfunctions. *Braz. J. Pain.* 2018; 1: 60–65.
 43. Yudkin J.S, Kumari M, Humphries S.E, Mohamed-Ali V. Inflammation, obesity, stress and coronary heart disease: Is interleukin-6 the link? *Atherosclerosis* 2000; 148: 209–214
 44. Tedgui A, Mallat Z. Cytokines in atherosclerosis: Pathogenic and regulatory pathways. *Physiol. Rev.* 2006;86: 515–581.
 45. Gunson M.J, Arnett G.W, Milam S.B. Pathophysiology and pharmacologic control of osseous mandibular condylar resorption. *J. Oral Maxillofac. Surg.* 2012; 70: 1918–1934.
 46. Kaplanski G, Marin V, Montero-Julian F, Mantovani A, Farnarier C. IL-6: A regulator of the transition from neutrophil to monocyte recruitment during inflammation. *Trends Immunol.* 2003; 24: 25–29.
 47. de Alcântara Camejo F, Azevedo M, Ambros V, Caporal K.S.T, Doetzer A.D, Almeida L.E. Interleukin-6 expression in disc derangement of human temporomandibular joint and association with osteoarthritis. *J. Cranio-Maxillofac. Surg.* 2017; 45: 768–774.



48. Hedges J.C, Singer C.A, Gerthoffer W.T. Mitogen-activated protein kinases regulate cytokine gene expression in human airway myocytes. *Am. J. Respir. Cell Mol. Biol.* 2000; 23: 86–94.
49. Sato J, Segami N, Kaneyama K, Yoshimura H, Fujimura K, Yoshitake Y. Relationship of calcitonin gene-related peptide in synovial tissues and temporomandibular joint pain in humans. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* 2004; 98: 533–540.
50. Koch A, Kunkel S, Burrows J, Evanoff H, Haines G, Pope R.M, et-al. Synovial tissue macrophage as a source of the chemotactic cytokine IL-8. *J. Immunol.* 1991;14: 2187–2195.
51. Koch A.E, Polverini P.J, Kunkel S.L, Harlow L.A, DiPietro L.A, Elner V.M, et-al. Interleukin-8 as a macrophage-derived mediator of angiogenesis. *Science* 1992; 258: 1798–1801.
52. Hirani N, Antonicelli F, Strieter R.M, Wiesener M.S, Ratcliffe P.J, Haslett C, et-al. The regulation of interleukin-8 by hypoxia in human macrophages—A potential role in the pathogenesis of the acute respiratory distress syndrome (ARDS). *Mol. Med.* 2001; 7: 685–697.
53. Nishimura M, Segami N, Kaneyama K, Suzuki T, Miyamaru M. Proinflammatory cytokines and arthroscopic findings of patients with internal derangement and osteoarthritis of the temporomandibular joint. *Br. J. Oral Maxillofac. Surg.* 2002; 40:68–71.
54. Sato J, Segami N, Nishimura M, Yoshitake Y, Kaneyama K, Kitagawa Y. Expression of interleukin 8 in synovial tissues in patients with internal derangement of the temporomandibular joint and its relationship with clinical variables. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* 2007; 103:467–474.
55. Murphy P.M, Tiffany H.L. Cloning of complementary DNA encoding a functional human interleukin-8 receptor. *Science* 1991; 253: 1280–1283
56. Schulte W, Bernhagen J, Bucala R. Cytokines in sepsis: Potent immunoregulators and potential therapeutic targets—An updated view. *Mediat. Inflamm.* 2013; 165-974.
57. Black R.A, Rauch C.T, Kozlosky C.J, Peschon J.J, Slack J.L, Wolfson M.F, et al. A metalloproteinase disintegrin that releases tumour-necrosis factor- α from cells. *Nature.* 1997;385: 729–733.
58. Kriegler M, Perez C, DeFay K, Albert I, Lu S.D. A novel form of TNF/cachectin is a cell surface cytotoxic transmembrane protein: Ramifications for the complex physiology of TNF *Cell.* 1988; 53: 45–53.
59. Cawthorn W.P, Sethi J.K. TNF- α and adipocyte biology. *FEBS Lett.* 2008;582: 117–131.
60. Bemelmans M, van Tits L, Buurman W. Tumor necrosis factor: Function, release and clearance *Crit. Rev. Immunol.* 1996; 16: 1–11.
61. Vanamee É.S, Faustman D.L. Structural principles of tumor necrosis factor superfamily signalling. *Sci. Signal.* 2018; 2(11):511
62. Wojdasiewicz P, Poniatowski Ł.A, Szukiewicz D. The role of inflammatory and anti-inflammatory cytokines in the pathogenesis of osteoarthritis. *Mediators Inflamm.* 2014; 2014: 561459.
63. Bazzoni F, Beutler B. The tumor necrosis factor ligand and receptor families. *N. Engl. J. Med.* 1996; 334:1717–1725.
64. Ahmed N, Catrina A.I, Alyamani A.O, Mustafa H, Alstergren P. Deficient cytokine control modulates temporomandibular joint pain in rheumatoid arthritis. *Eur. J. Oral Sci.* 2015; 123: 235–241.
65. Fredriksson L, Alstergren P, Kopp S. Tumor necrosis factor- α in temporomandibular joint synovial fluid predicts treatment effects on pain by intra-articular glucocorticoid treatment. *Mediat. Inflamm.* 2006(6), 59425
66. Kringel D et al. A machine-learned analysis of human gene polymorphisms modulating persisting pain points to major roles of neuroimmune processes. *Eur. J. Pain.* 2018; 22: 1735–1756
67. Puliti M. et al. Contribution of matrix metalloproteinase 2 to joint destruction in group B streptococcus-induced murine arthritis. *Arthritis Rheumatol.* 2012; 64:1089–1097 .
68. Yoshida K. et al. Expression of matrix metalloproteinases and aggrecanase in the synovial fluids of patients with symptomatic temporomandibular disorders. *Oral. Surg. Oral. Med. Oral. Pathol. Oral. Radio. Endod.* 2006;102; 22–27.
69. Tabeian H. et al. IL-1 β damages brocartilage and upregulates MMP-13 expression in brochondrocytes in the condyle of the temporomandibular joint. *Int J. Mol. Sci.* 2019; 20: 2260.
70. Gho W.G, Choi Y, Park K. H. & Huh, J. K. Expression of collagenases (matrix metalloproteinase-1, 8, 13) and tissue inhibitor of metalloproteinase-1 of ret-rodiscal tissue in temporomandibular joint disorder patients. *J. Korean Assoc. Oral. Maxillofac. Surg.* 2018; 44: 120–127.
71. Loreto C. et al. MMP-7 and MMP-9 are overexpressed in the synovial tissue from severe temporomandibular joint dysfunction. *Eur. J. Histochem* 2020; 64: 3113 .
72. Srinivas R. et al. Matrix metalloproteinases in mild and severe tempor-omandibular joint internal derangement synovial uid. *Oral. Surg. Oral. Med Oral. Pathol. Oral. Radio. Endod.* 2001; 91: 517–525 .
73. Kanyama M. et al. Matrix metalloproteinases and tissue inhibitors of metallo-proteinases in synovial uids of patients with temporomandibular joint osteoarthritis. *J. Orofac. Pain.* 2000; 14: 20–30 .
74. Fujita H. et al. MMP-3 activation is a hallmark indicating an early change in TMJ disorders and is related to nitration. *Int J. Oral. Maxillofac. Surg.* 2009; 38: 70–78 .
75. Huang X. et al. Effect of psychological stress on the structure of the tempor-omandibular joint and the expression of MMP-3 and TIMP-3 in the cartilage in rats. *Br. J. Oral. Maxillofac. Surg.* 2014; 52: 709–714 .
76. Nascimento G.C, Rizzi E, Gerlach R. F. & Leite-Panissi, C. R. Expression of MMP-2 and MMP-9 in the rat trigeminal ganglion during the development of tem- poromandibular joint in ammation. *Braz. J. Med Biol. Res.* 2013; 46: 956–967 .
77. Nishimura M, Segami N, Kaneyama, K., Suzuki, T. & Miyamaru, M. Relationships between pain-related mediators and both synovitis and joint pain in patients with internal derangements and osteoarthritis of the temporomandibular joint. *Oral. Surg. Oral. Med Oral. Pathol. Oral. Radio. Endod.* 2002; 94: 328–332.
78. Miller, K. E., Hoffman, E. M., Sutharshan, M. & Schechter, R. Glutamate pharmacology and metabolism in peripheral primary afferents: physiological and pathophysiological mechanisms. *Pharm.* 2011; 130: 283–309.
79. Lam D. K, Sessle B. J, Cairns B. E. & Hu, J. W. Neural mechanisms of tempor omandibular joint and masticatory muscle pain: a possible role for peripheral glutamate receptor mechanisms. *Pain. Res. Manag.* 2005; 10: 145–152.
80. Alstergren, P. et al. Glutamate-induced temporomandibular joint pain in healthy individuals is partially mediated by peripheral NMDA receptors. *J. Orofac. Pain.* 2010; 24: 172–180 .
81. Sommer C. Is serotonin hyperalgesic or analgesic? *Curr. Pain. Headache Rep.* 2006; 10:101–106).
82. Dawson A, Stensson N, Ghafouri B, Gerdle B, List T, Svensson P, et al. Dopamine in plasma – a biomarker for myofascial TMD pain? *The Journal of Headache and Pain.* 2018 Oct 15:17(1).
83. Wood P, Patterson J, Sunderland J. Reduced presynaptic dopamine activity in fibromyalgia syndrome demonstrated with positron emission tomography: a pilot study. *J Pain.* 2007;8(1):51–8.
84. Jasim H, Louca S, Christidis N. Salivary cortisol and psychological factors in women with chronic and acute oro-facial pain. *J Oral Rehabil.* 2014;41(2):122–32.
85. Maixner W, Greenspan J, Dubner R. Potential autonomic risk factors for chronic TMD: descriptive data and empirically identified domains from the OPPERA case- control study. *J Pain.* 2011;12(11):75–91.
86. Ernberg M. in *Peripheral Receptor Targets for Analgesia* (eds Cairns, B. E. 2009;243–274 (John Wiley & Sons, Inc.).
87. Ernberg M. et al. Pain allodynia and serum serotonin level in orofacial pain of muscular origin. *J. Orofac. Pain.* 1999; 13: 56–62 .
88. Kopp S. Influence of neuropeptides, serotonin and interleukin1 beta on tem poromandibular joint pain and in ammation. *J. Oral. Maxillofac. Surg.* 1998; 56: 189–191 .



89. Ernberg M. et al. Plasma and serotonin levels and their relationship to orofacial pain and anxiety in bromyalgia. *J. Orofac. Pain.* 2000; 14; 37–46.
90. Kopp S& Alstergren, P. Blood serotonin and joint pain in seropositive versus seronegative rheumatoid arthritis. *Mediators In amm.* 2002, 11, 211–217.
91. Fredriksson, L., Alstergren, P. & Kopp, S. Serotonergic mechanisms in uence the response to glucocorticoid treatment in TMJ arthritis. *Mediators In amm.* 2005, 194–201
92. Ali SQ, Hadi R. Assessment of Cortisol as Salivary Psychological Stress Marker in Relation to Temporomandibular Disorders among a Sample of Dental Students. *Journal of Baghdad College of Dentistry.* 2015;27(2):86–92.
93. Hoehn K, Marieb E. *Human Anatomy and Physiology.* San Francisco: Benjamin Cummings; 2010.
94. Barbosa T, Castelo P, Leme M, Gavião M. Associations between oral health-related quality of life and emotional statuses in children and preadolescents: Oral health-related quality of life and emotional statuses. *Oral Diseases.* 2012 Oct;18(7):639–47.
95. Benedetta C. Salivary cortisol levels in women with muscular facial pain.
96. AlSahman, L.; AlBagieh, H.; AlSahman, R.; Mokeem, N.R.; Costa, L.P. Does salivary cortisol serve as a potential biomarker for temporomandibular disorders in adults? *BMC Oral Health* 2024; 24: 1364.
97. Wen S, Santander J, Barria D, Salazar LA, Sandoval C, Arias C. Epigenetic Biomarkers in Temporomandibular Joint Osteoarthritis: An Emerging Target in Treatment. *Int J Mol Sci.* 2025; 26(8):3668.
98. Thamer, S.R.; Diajil, A.R. Therapeutic effect of intra-articular injection of hyaluronic acid and platelet-rich plasma on the expression of salivary matrix metalloproteinases 2 and 9 in tempromandibular internal derrangement patients. *J. Emerg. Med. Trauma Acute Care* 2024; 2024: 3
99. AlSahman L, AlBagieh H, AlSahman R, Mokeem N.R, Costa L.P. Does salivary cortisol serve as a potential biomarker for temporomandibular disorders in adults? *BMC Oral Health* .2024; 24: 1364.
100. Ismah N, Bachtiar E, Purwanegara M, Tanti I, Mardiaty E, Ismah N, et-al. Evaluation of IL-1 β and CRP mRNA expression levels by RT-PCR in postorthodontic treatment patients with temporomandibular joint disorders: A cross-sectional Study. *J. Int. Soc. Prev. Community Dent.* 2024; 14: 98–104.
101. Kim Y, Son C, Park Y.K, Jo J.H, Park J.W. Sleep duration and inflammatory mediator levels associated with long-term prognosis in temporomandibular disorders. *J. Oral Rehabil.* 2023; 50: 830–839.
102. Ornek Akdogan E, Omezli MM, Torul D. Comparative evaluation of the trabecular structure of the mandibular condyle, the levels of salivary cortisol, MMP-3, TNF- α , IL- 1 β in individuals with and without temporomandibular joint disorder. *J Stomatol Oral Maxillofac Surg.* 2023;124:101417.
103. Shao B., Xu Y, Jia M, Li C, Gong Z, Shao B, Xu Y, Jia M, Li C.-x.; Gong, Z.-c. Association of HMGB1 levels in synovial fluid with the severity of temporomandibular joint osteoarthritis. *BMC Musculoskelet. Disord.* 2023;24
104. Aparna N, Rajesh S, Ranukumari A, Shakila R. A case-control investigation of the psychological and physiological stress markers with salivary cortisol levels in patients with temporomandibular joint disorders: A short clinical study. *J. Indian Prosthodont. Soc.*2023; 23: 163–169
105. Kazan D, Burcu B.A, Akzoy A, Enes A. The evaluation of oxidative stress and inflammation markers in serum and saliva of the patients with temporomandibular disorders. *Turk. J. Med. Sci.* 2023; 53: 1690–1696.
106. Zwiri A.M, Al-Hatamleh M.A.I, Al-Wakeel W.M.A, Asif J.A, Khoo S.P, Husein A, et-al. A Randomized Controlled Trial Evaluating the Levels of the Biomarkers hs-CRP, IL-6, and IL-8 in Patients with Temporomandibular Disorder Treated with LLLT, Traditional Conservative Treatment, and a Combination of Both. *Int. J. Environ. Res. Public Health.* 2022; 19: 8987.
107. Ulmner M, Sugars R, Naimi-Akbar A, Alstergren P, Lund B, Ulmner M, et-al. Cytokines in temporomandibular joint synovial fluid and tissue in relation to inflammation. *J. Oral Rehabil.* 2022; 49:599–607.
108. Bayindir S, Asan C, Demirbas A, Ketik D, Kütük N, Bayindir S, et-al. Evaluation of aggrecan and adipokine levels in temporomandibular joint synovial fluid. *J. Cranio-Maxillofac. Surg.* 2022; 50: 432–438.
109. Venkatesh S.B, Shetty S.S, Kamath V. Prevalence of temporomandibular disorders and its correlation with stress and salivary cortisol levels among students. *Pesqui. Bras. Odontopediatria Clin. Integr.* 2021; 21: 1–9.
110. Son C, ParkY.K, Park J.W. Long-term evaluation of temporomandibular disorders in association with cytokine and autoantibody status in young women. *Cytokine* 2021;144: 155551.
111. Ulmner M, Sugars R, Naimi-Akbar A, Suslu S, Reseland J, Kruger-Weiner C, et al. Synovial tissue cytokine profile in disc displacement of the temporomandibular joint. *J. Oral Rehabil.* 2020; 47: 1202–1211
112. Loreto C, Filetti V, Almeida L.E, Rapisarda G.R.M, La Rocca R, Graziano C, et-al. MMP-7 and MMP-9 are overexpressed in the synovial tissue from severe temporomandibular joint dysfunction. *Eur. J. Histochem. EIJH* 2020; 64: 3113.
113. Khayamzadeh M, Mirzaei Dizgah I, Aghababainejad P, Habibzadeh S, Kharazifard MJ. Relationship between parafunctional habits and salivary biomarkers. *Front Dent.* 2020;16:465- 472.
114. Shoukri B, Prieto JC, Ruellas A, et al. Minimally invasive approach for diagnosing TMJ osteoarthritis. *J Dent Res.*2019;98:1103- 1111.
115. Xiong H, Li W, Li J, Fang W, Ke J, Li B, et-al. Elevated leptin levels in temporomandibular joint osteoarthritis promote proinflammatory cytokine IL-6 expression in synovial fibroblasts. *J. Oral Pathol. Med.* 2019; 48: 251–259.
116. Yang M.C, Wang D.H, Wu H.T, Li W.C, Chang T.Y, Lo W.L, et-al. Correlation of magnetic resonance imaging grades with cytokine levels of synovial fluid of patients with temporomandibular joint disorders: A cross-sectional study. *Clin. Oral Investig.* 2019; 23: 3871–3878.
117. Ganti S, Sukhija P, Ahmad A.S, Kumar J.M, Ali A, Dhir A. Evaluation of Effect of Glucosamine-Chondroitin Sulfate, Tramadol, and Sodium Hyaluronic Acid on Expression of Cytokine Levels in Internal Derangement of Temporomandibular Joint. *J. Contemp. Dent. Pract.* 2018; 19: 1501–1505.
118. Cê P, Barreiro B, Silva R, Oliveira R, Heitz C, Campos M. Salivary levels of Interleukin- 1 α in temporomandibular disorders and fibromyalgia. *J Oral Facial Pain Headache.* 2018;32:130- 136.
119. Watanabe S, Ogura N, Akutsu M, Kawashima M, Hattori T, Yano T, et-al. Pro-inflammatory Cytokine Production in Co-culture of Human Monocytes and Synovial Fibroblasts from the Human Temporomandibular Joint. *Int. J. Oral—Med. Sci.* 2017; 15: 107–113.
120. Kobayashi FY, Gavião MBD, Marquês MCS, et al. Salivary stress biomarkers and anxiety symptoms in children with and without temporomandibular disorders. *Braz Oral Res.* 2017;31:e78.
121. Dawson A. et al. Dopamine in plasma- a biomarker for myofascial TMD pain? *J. Headache Pain.* 2016; 17: 65.
122. Ahmed N, Catrina A.I, Alyamani A.O, Mustafa H, Alstergren P. Deficient cytokine control modulates temporomandibular joint pain in rheumatoid arthritis. *Eur. J. Oral Sci.* 2015; 123: 235–241
123. Lin S.L, Wang S.L, Tu C.C, Kao S.Y, Yeh J.W. Serum cortisol level and disc displacement disorders of the temporomandibular joint. *J. Oral Rehabil.* 2015; 43: 10–15.
124. Cevindanes LH, Walker D, Schilling J, Sugai J, Giannobile W, Paniagua B, et-al. 3D osteoarthritic changes in TMJ condylar morphology correlates with specific systemic and local biomarkers of disease. *Osteoarthritis Cartilage.* 2014 Oct;22(10):1657-67.
125. Vos L.M, Kallik R, Heffez J.J, Schimming B. Alteration of cartilage degeneration and inflammation markers in temporo



- mandibular joint osteoarthritis occurs proportionally. *J. Oral Maxillofac. Surg.* 2013; 71: 1659–1664.
126. Wake M, Hamada Y, Kumagai K, Tanaka N, Ikeda Y, Nakatani Y, et al. Up-regulation of interleukin-6 and vascular endothelial growth factor-A in the synovial fluid of temporomandibular joints affected by synovial chondromatosis. *Br. J. Oral Maxillofac. Surg.* 2013; 5: 164–169
 127. Nogami S, Takahashi T, Ariyoshi W, Yoshiga D, Morimoto Y, Yamauchi K. Increased levels of interleukin-6 in synovial lavage fluid from patients with mandibular condyle fractures: Correlation with magnetic resonance evidence of joint effusion. *J. Oral Maxillofac. Surg.* 2013; 71: 1050–1058.
 128. Kim Y.K, Kim S.G, Kim B.S, Lee J.Y, Yun P.Y, Bae J.H, et al. Analysis of the cytokine profiles of the synovial fluid in a normal temporomandibular joint: Preliminary study. *J. Cranio-Maxillofac. Surg.* 2012; 40: e337–e341
 129. Slade G.D, Conrad M.S, Diatchenko L, Rashid N.U, Zhong S, et al. Cytokine biomarkers and chronic pain: Association of genes, transcription, and circulating proteins with temporomandibular disorders and widespread palpation tenderness. *Pain* 2011;152: 2802–2812.
 130. Lee J.K, Cho Y.S, Song S.I. Relationship of synovial tumor necrosis factor alpha and interleukin 6 to temporomandibular disorder. *J. Oral Maxillofac. Surg.* 2010; 68: 1064–1068. *Diagnostics* 2020; 10:303 14 of 18
 131. Hamada Y, Kondoh T, Holmlund A.B, Sakota K, Nomura Y, Seto K. Cytokine and clinical predictors for treatment outcome of visually guided temporomandibular joint irrigation in patients with chronic closed lock. *J. Oral Maxillofac. Surg.* 2008; 66: 29–34
 132. Vernal R, Velasquez E, Gamonal J, Garcia-Sanz J.A, Silva A, Sanz M. Expression of proinflammatory cytokines in osteoarthritis of the temporomandibular joint. *Arch. Oral Biol.* 2008; 53: 910–915
 133. Kardel R, Ulfgren A.K, Reinholt F, Hamada Y, Holmlund A. Inflammatory cell and cytokine patterns in patients with chronic polyarthritis and temporomandibular joint involvement. *Acta Odontol. Scand.* 2006; 64: 221–226.
 134. Matsumoto K, Honda K, Ohshima M, Yamaguchi Y, Nakajima I, Micke P, et al. Cytokine profile in synovial fluid from patients with internal derangement of the temporomandibular joint: A preliminary study. *Dentomaxillofac. Radiol.* 2006; 35: 432–441
 135. Kaneyama K, Segami N, Suzuki J, Nishimura M, Hosaka Y. Interleukin-6 family of cytokines as biochemical markers of osseous changes in the temporomandibular joint disorders. *Br. J. Oral Maxillofac. Surg.* 2004; 42: 246–250
 136. Nishimura M, Segami N, Kaneyama K, Sato J, Fujimura K. Comparison of cytokine level in synovial fluid between successful and unsuccessful cases in arthrocentesis of the temporomandibular joint. *J. Oral Maxillofac. Surg.* 2004; 62: 284–287.
 137. Kardel R, Ulfgren A.K, Reinholt F, Holmlund A. Inflammatory cell and cytokine patterns in patients with painful clicking and osteoarthritis in the temporomandibular joint. *Int. J. Oral Maxillofac. Surg.* 2003; 32: 390–396
 138. Tanaka A, Kumagai S.S, Takatsuka S, Nakagawa K, Yamamoto E. Expression of matrix metalloproteinase-2 and -9 in synovial fluid of the temporomandibular joint accompanied by anterior disc displacement *J Oral Pathol Med* 2001; 30: 59–64
 139. Shinoda C, Takaku S. Interleukin-1 beta, interleukin-6, and tissue inhibitor of metalloproteinase-1 in the synovial fluid of the temporomandibular joint with respect to cartilage destruction. *Oral Dis.* 2000; 6: 383–390.
 140. Murakami K.I, Shibata T, Kubota E, Maeda H. Intra-articular levels of prostaglandin E2, hyaluronic acid, and chondroitin-4 and -6 sulfates in the temporomandibular joint synovial fluid of patients with internal derangement. *J. Oral Maxillofac. Surg.* 1998; 56: 199–203
 141. Kubota E, Kamata T, Murakami J, Shibata T, Murakami K.I. Synovial fluid cytokines and proteinases as markers of temporomandibular joint disease. *J. Oral. Maxillofac. Surg.* 1998; 56: 192–198.
 142. Takahashi T, Kondoh T, Fukuda M, Yamazaki Y, Toyosaki T, Suzuki R. Proinflammatory cytokines detectable in synovial fluids from patients with temporomandibular disorders. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* 1998; 85:135–141
 143. Kubota E, Horiuchi I, Kamata T, Shibata T, Murakami K. Interleukin 1 beta and stromelysin (MMP3) activity of synovial fluid as possible markers of osteoarthritis in the temporomandibular joint. *J. Oral Maxillofac. Surg.* 1997; 55: 20–27.
 144. Fu K, Ma X, Zhang Z, Pang X, Chen W. Interleukin-6 in synovial fluid and HLA-DR expression in synovium from patients with temporomandibular disorders. *J. Orofac. Pain* 1995; 9: 131–137.